



Trade Science Inc.

January 2007

Volume 1 Issue 1

BioCHEMISTRY

An Indian Journal

Regular Paper

BCAIJ, 1(1), 2007 [8-13]

Simple And Rapid Methods For The Analysis Of Captopril In Dosage Form



A.C.Sharada
Department of Biochemistry,
Yuvaraja's College,
University of Mysore,
Mysore-570005 (INDIA)
Ph- 984567919
E-mail. sharadaac@gmail.com

Received: 25th September, 2006

Accepted: 7th November, 2006

Web Publication Date : 21st December, 2006



H.Chandru
Department of Studies in Biochemistry,
University of Mysore,
Manasagangotri, Mysore, (INDIA)

ABSTRACT

Two indirect methods are described for the micro determination of captopril using hexacyanoferrate (III) as reagent. Titrimetry involves the addition of a known excess of hexacyanoferrate (III) to the sample solution and back titration of the residual reagent iodometrically in the presence of a large excess of zinc sulphate, after the specified time. In spectrophotometry, the sample solution is treated with a large excess of hexacyanoferrate (III) and after specified time, hexacyanoferrate (II), the reduced form of the reagent was reacted with orthophenanthroline, and the absorbance of the resulting red colour measured at 510 nm, which forms the basis for the quantitative determination of captopril. The reaction used for titrimetry proceeds at room temperature and will be complete in 10 minute with a stoichiometry of 1:1 with respect to the oxidant and captopril. The reaction product used for spectrophotometric determination shows the absorption maximum at 510 nm. The Beer's law is obeyed over the concentration range 0.25-12.00 $\mu\text{g ml}^{-1}$, the molar absorptivity and Sandell sensitivity for the system being $9.14 \times 10^3 \text{ l mol}^{-1} \text{ cm}^{-1}$ and 23.78 ng cm^{-2} , respectively. The limit of detection and quantification are found to be 0.08 and 0.26 $\mu\text{g ml}^{-1}$, respectively. Both procedures have been applied to the determination of captopril in tablets. The results have been statistically compared with those obtained by the official (BP) method.

© 2007 Trade Science Inc. - INDIA

KEYWORDS

Captopril;
Hexacyanoferrate;
Titrimetry;
Spectrophotometry;
Oxidation reaction.

INTRODUCTION

Captopril, 1-(3-mercapto-2-D-methyl-1-oxopropyl)-1-proline, is an antihypertensive drug widely prescribed in the treatment of hypertension and chronic congestive heart failure. It acts primarily by inhibiting the angiotensin-converting enzyme. Simple methods for its assay in technical and formulation grade sample are necessary for routine analysis and quality evaluation. The most widely used technique for the determination of captopril is high performance liquid chromatography^[1-5] but the procedures based on this technique require either derivatisation of the compound or selective detectors and elaborate multistep extraction. Gas chromatographic methods reported either involve derivatisation^[6,7] or lack the required sensitivity^[8] or restricted to body fluids^[9-12]. Several other instrumental methods such as differential polarography^[13], capillary electrophoresis^[14], spectrofluorimetry^[15], amperometry^[16] and coulometry^[17] have been applied for the assay of drugs in pharmaceuticals. Quantification of captopril has also been achieved by radio^[18] and enzyme^[19] immunoassay methods. Literature on the titrimetric methods for the analysis of captopril is scanty. Three methods^[20-22] reported use silver nitrate as the reagent.

Direct spectrophotometric methods based on diazotization and coupling reaction^[23], redox reaction using Folin-Cioltu reagent^[24] and molybdophosphoric acid^[25] and complex formation reaction involving dithiopyridine^[26], phenylfurazone^[27] and palladium (II) chloride^[28] have been used for the determination of the drug. The procedures suffer from one or the other disadvantage such as insufficient sensitivity^[23] (5-20mgml⁻¹), selectivity^[24,25] or longer contact time^[27] (1h). Indirect spectrophotometric methods involving the measurement of decrease in absorbance cresyl fast violet acetate^[29], celestine blue^[30] and 2,2-diphenyl-1-picryl hydrazyl^[31] have also been reported for the assay captopril. However, the accuracy and reliability of these methods are heavily dependent on the purity of the coloured reagents. In search of more convenient methods, titrimetric and spectrophotometric investigations of the oxidation reaction between captopril and hexacyanoferrate (III) have been carried out and their application to the analysis

of authentic samples and dosage forms have been examined.

EXPERIMENTAL

Apparatus

A Systronics model 106 digital spectrophotometer with 1 cm matched glass cells was used for absorbance measurements.

Reagents and materials

All chemicals used were of analytical reagent grade and distilled water was used to prepare all solutions. A standard solution of 0.02M potassium hexacyanoferrate (III) was prepared by dissolving 6.5852 g of reagent in one liter of water in a volumetric flask. Sodium thiosulphate solution (~0.02M) was prepared in water and standardized iodometrically^[32]. Zinc sulphate (30%), potassium iodide (10%), hydrochloric acid (5M) and starch indicator (1%) solutions were prepared in the usual way and used in titrimetric work. Hexacyanoferrate (III) (100 µg ml⁻¹) obtained by stepwise dilution of 0.02M solution, 0.25% 1,10-phenanthroline, 1.5 M hydrochloric acid and 0.5M sodium carbonate solutions were employed for spectrophotometric investigation.

Titrimetry

To a 10 ml aliquot containing 1-8 mg of captopril in a titration flask were added 2 ml of 5M hydrochloric acid, 10 ml of 0.02M hexacyanoferrate (III) and 10ml of 30% zinc sulphate solutions, mixed well and kept aside for 20 minute with occasional shaking. Then, 5 ml of 10% potassium iodide solution were added and the liberated iodine was titrated with 0.02M thiosulphate solution in the presence of starch indicator. A blank experiment was carried out in the same manner. Amount of drug was calculated from the equation:

$$\text{Drug (mg)} = (V_1 - V_2) MR/n$$

Where V_1 = Volume of thiosulphate consumed in the blank titration, ml;

V_2 = Volume of thiosulphate consumed in the sample titration, ml;

M = Relative molar mass of drug;

Regular Paper

R = Molarity of hexacyanoferrate (III)
and n = Number of moles hexacyanoferrate (III) reacting per mole of drug.

Spectrophotometry

A 3.0ml aliquot of standard drug solution containing 2.5-120 μg of captopril was placed in a 10 ml volumetric flask to which 1.5 ml of 5 M hydrochloric acid and 1ml of 100 $\mu\text{g ml}^{-1}$ of hexacyanoferrate (III) solutions were added. The flasks and contents were swirled, placed in a boiling water bath for 10 minute. Then, the flasks were cooled, 1.5 ml of 0.5M sodium carbonate were added followed by 1ml of 0.25% 1,10-phenanthroline and diluted to mark with water. The coloured product was measured at 510nm against the reagent blank. The concentration of drug in the unknown was deduced from the calibration graph or computed from the regression equation.

Tablets

Twenty tablets were weighed and ground into a fine powder. A quantity of powder containing 100 mg of active ingredient was weighed accurately into a 100 ml volumetric flask, 60 ml of water added and shaken thoroughly for about 20 minute to extract the drug. Diluted to the mark with water, mixed well and filtered through a Whatman No. 41 filter paper. First ten of the filtrate was rejected. A convenient volume was assayed by titrimetry using the general procedure. The filtrate was diluted appropriately to get 40 $\mu\text{g ml}^{-1}$ and a suitable volume was subjected to analysis by spectrophotometry.

RESULTS AND DISCUSSION

Both methods are based on the oxidation of captopril with excess of hexacyanoferrate (III). In titrimetry, the excess oxidant, after oxidation was determined iodometrically in the presence of zinc sulphate, there by allowing indirect assay of captopril, whereas in spectrophotometry, hexacyanoferrate (II), a reaction product is determined by reacting it with 1,10-phenanthroline, thus permitting the indirect estimation of captopril.

The reaction variables were optimized by varying each variable while keeping others constants for

obtaining constant stoichiometry in titrimetry and reproducible and maximum absorbance in spectrophotometry. The oxidation reaction was found to be quantitative in hydrochloric acid medium and 2 ml of 5M acid in a total volume 25 ml was found to produce a constant molar ratio. In the back titration of hexacyanoferrate (III) iodometrically, zinc sulphate was added to ensure quick and irreversible oxidation of iodide by hexacyanoferrate (III)^[33] through the removal of hexacyanoferrate (II) forme^[34] as the slightly soluble potassium zinc hexacyanoferrate (II). The oxidation of the drug by hexacyanoferrate (III) was slow at room temperature and found to be complete in 20 minute under specified acid and oxidant concentrations. Two fold increases in excess of hexacyanoferrate (III) had no effect on the stoichiometry of the reaction. For the range studied (1-8mg), a 10ml volume of 0.02M hexacyanoferrate (III) gave stoichiometric results. Under the stated experimental conditions, 2 moles of hexacyanoferrate (III) were consumed per mole of captopril. The linearity between the amount of the drug and the titration end point is apparent from the value of the correlation coefficient, r (0.9982) obtained by the best fit-line via linear least squares treatment.

Spectrophotometric method is based on the oxidation reaction of captopril with hexacyanoferrate (III) under acidic conditions and subsequent determination of hexacyanoferrate (II), the reduced form of the oxidant, by reacting with 1,10-phenanthroline under mildly acidic conditions (pH 3-4) resulting in the formation of red colour^[35] which could be measured at 510nm. Captopril when added in increasing amounts to a fixed amount of hexacyanoferrate (III), there is a concomitant rise in the concentration of hexacyanoferrate (II), after the oxidation reaction. This is observed as a proportional increase in the absorbance of the reaction mixture on adding 1,10-phenanthroline. The oxidation reaction was slow at room temperature and was found to be complete when heated with 5 M hydrochloric acid in a boiling water-bath for 10 minutes. For the range investigated (2.5-120 μg), 1ml of 100 $\mu\text{g ml}^{-1}$ hexacyanoferrate (III) and 1.5 ml of 5M hydrochloric acid were sufficient to effect oxidation. Since the colour reaction between

hexacyanoferrate (II) and 1,10-phenanthroline takes place in the pH range 3-4, 1.5 ml of 0.5M sodium carbonate solution were needed to raise the pH to about 3.5 before adding 1,10 phenanthroline solution. The colour produced was found to be stable for several weeks.

Under the experimental conditions described, Beer's law is obeyed over the concentration range 0.25-12 $\mu\text{g ml}^{-1}$. The linear regression equation:

$$A = 0.01 + 0.04 C \quad (r = 0.9982)$$

Where A is the absorbance and C is the concentration in $\mu\text{g ml}^{-1}$. The apparent molar absorptivity was $9.14 \times 10^3 \text{ l mol}^{-1} \text{ cm}^{-1}$ and Sandell sensitivity was 23.78 ng cm^{-2} . The limit of detection was $0.0776 \mu\text{g ml}^{-1}$ and the limit of quantification as the lowest standard concentration, which could be determined with acceptable accuracy, and precision was $0.2589 \mu\text{g ml}^{-1}$.

Accuracy and precision

The accuracy and precision of the methods were checked by seven replicate determinations at 2, 5 and 7 mg levels by titrimetry and 30, 60 and 90 μg levels by spectrophotometry. The percent error was found to be less than 2 and the relative standard deviation

found to be between 0.45 and 2.13, which are indicative of satisfactory accuracy and repeatability. The results of the study are compiled in TABLE 1.

Application

The results for the determination of captopril in the three brands of tablets are given in TABLE 2. In which they are compared with those obtained using reported established methods^[35]. Statistical analysis of the results by the variance-ratio test and t-test showed that there was no significant difference between the performance of the proposed method and reference methods in terms of reproducibility and accuracy.

The accuracy and validity of the methods were further assessed by performing recovery studies. When tablets preparations spiked with known amounts pure drug were analysed, quantitative recoveries (99.98%) were obtained indicating that lactose, talc, starch, magnesium stearate and sodium alginate, which are commonly incorporated as excipients did not interfere.

TABLE 1: Evaluation of accuracy and precision

Titrimetry				Spectrophotometry			
Amount taken, Mg	Amount found*, Mg	Error, %	RSD, %	Amount taken, μg	Amount found, μg	Error, %	RSD, %
2.00	2.04	2.0	2.13	30.00	29.23	0.23	0.93
5.00	5.02	0.40	1.81	60.00	60.11	0.18	0.45
7.00	7.06	0.86	1.32	90.07	90.07	0.07	0.45

*Average value of seven determinations
RSD- Relative standard deviation

TABLE 2: Results of analysis of tablets using the proposed methods

Tablet brand name*	Label claim mg per tablet	Found ϕ (% recovery \pm SD)			t-value [#]		F-value [@]	
		Titrimetry (T)	Spectro photometry (S)	Reference methods	T	S	T	S
Acetin ^{α}	25.00	99.14 \pm 0.38	97.96 \pm 0.85	99.76 \pm 0.62	1.84	3.60	2.66	1.88
Angiopril ^{β}	25.00	100.28 \pm 0.64	102.23 \pm 1.02	100.62 \pm 0.46	0.92	3.24	1.94	4.90
	50.00	98.10 \pm 0.36	98.76 \pm 0.67	97.56 \pm 0.38	2.17	3.37	1.11	3.11
Captopril ^{γ}	12.50	101.28 \pm 0.53	101.66 \pm 0.96	100.84 \pm 0.56	1.20	1.61	1.12	2.94
	25.00	98.63 \pm 0.66	99.35 \pm 0.74	98.78 \pm 0.26	0.48	1.70	6.44	8.10

Ψ Mean value of five determinations, [#] Tabulated value at 95% confidence level is 2.77, [@] Tabulated value at 95% confidence level is 6.39, * Marketed by: α -Wockhardt, β -Torrent, γ -Lupin Lab Ltd.

Regular Paper

TABLE 3: Results of recovery study using standard-addition method

Tablet brand name studied	Titrimetry				Spectrophotometry			
	Amount of drug in extract, mg	Amount of pure drug, mg	Total found, mg	Recovery* of pure drug added, %	Amount of drug in extract, µg	Amount of pure drug, µg	Total found, µg	Recovery* of pure drug added, %
Acetin (25 mg)	0.9914	3.00	3.934	98.08	19.59	30.00	48.96	97.90
	0.9914	5.00	5.850	97.17	19.59	50.00	70.54	101.90
	0.9914	7.00	7.79	97.12	19.59	80.00	102.16	103.21
Angiopril (25 mg)	1.0000	3.00	4.087	102.90	20.00	30.00	51.89	102.29
	1.0000	5.00	6.043	100.86	20.00	50.00	70.89	101.78
	1.0000	7.00	8.109	101.56	20.00	80.00	99.63	99.53
Captopril (25 mg)	0.9863	3.00	4.048	102.06	19.73	30.00	50.78	103.50
	0.9863	5.00	5.95	99.27	19.73	50.00	70.25	101.04
	0.9863	7.00	7.89	98.62	19.73	80.00	101.09	101.69

* Average value of three trials

CONCLUSION

The methods are simple, sensitive and their application to the determination of the micro and sub micro amounts of captopril has been demonstrated.

ACKNOWLEDGEMENT

The authors thank Torrent Pharmaceuticals, Ahmedabad, India, for gifting pure sample of captopril. We also thank to prof. M. Karunakumar Chairman Department of Biochemistry, University of Mysore to provided laboratory facilities to perform this work.

REFERENCES

- [1] A.Khedr, H.EL-Sherif; Biomed Chromatogr, **12**, 57 (1998).
- [2] J.Russel, J.A.Mc keown, C.Hensman, W.E.Smith, J.Raglans; J Pharm Anal, **15**, 1757 (1998).
- [3] E.Bald, S.Sypniewski, Fresenins; J Anal Chem, **358**, 554 (1997).
- [4] J.Wieling, G.Hendrikis, W.J.Tamminga, J.Hemepenius, C.K.Mensink, B.Oosterhuis, J.H.G.Jonkman; J.Chromatogr, **730**, 318 (1996).
- [5] V.Cavrini, R.Gatti, A.M.Dipietra, M.A.Raggi; Chromatographia, **23**, 680 (1987).
- [6] M.Jemal, A I.Cohen; Anal.Chem, **57**, 2407 (1985).
- [7] Y.C.Liu, H.L.Wu, H.S.Kou, S.H.Chen, S.M.Wu; Anal Lett, **28**, 1465 (1995).
- [8] C.H.Liu, S.L.Liu, H.N.Chen, X.T.Xie; Sepu, **16**, 62 (1998).
- [9] M.E.Franklin, R.S.Addson, P.U.Baker, W.D.Hooper; J Chromatogr Biomed Appl, **705**, 47 (1998)
- [10] H.J.Leis, M.Leis, W.Weiz, E.Malle; J.Chromatography Biomed Appl, **94**, 299 (1990).
- [11] T.Ito, Y.Matsuki, H.Kurihara, T.Nambara; J Chromatogr Biomed Appl, **61**, 79 (1987).
- [12] M.Jemal, E.Ivashkiy, A.I.Cohen; Biomed.Mass Spectrum, **12**, 664 (1985).
- [13] J.M.G.Fraga, A.I.J.Abizanda, F.J.Moreno, J.J.A .Leon; Talanta, **46**, 75 (1998).
- [14] J.Russel, D.L.Rabenstein; Anal.Biochem., **242**, 136 (1996).
- [15] R.Segarra Gueerreo, S.Sagrado Vives, J.Martinez Calatagn; Microchem.J., **43**, 176 (1991).
- [16] Z.Korieanac, T.Jovanovic, B.Stankovic; Pharmazie, **50**, 299 (1995).
- [17] K.Nikolic Kand Velasevic; Acta Pol.Pharm, **48**, 5 (1991).
- [18] J.I.Tu, J.Brennan, B.Stouffer, W.C.Ecklelman; Ther Drug Monitor, **12**, 404 (1990).
- [19] H.Kinoshita, R.Nakamaru, S.Tanaka, Y.Tohira, M.Sawada; J Pharm Sci, **75**, 71 (1986).
- [20] K.Nikolic, K.R.Velasevic; J Pharm Belg, **45**, 17 (1990).
- [21] K.Nikolic, K.R.Velasevic; Pharmzie, **44**, 155 (1989).
- [22] M.M.Buzanova, I.V.Kasaudi; Zavod Lab, **61**, 7 (1995).
- [23] N.M.Sanghavi, M.M.Samarth, R.S.P.S.Mathru; Indian Drugs, **28**, 489 (1991).
- [24] C.S.P.Sastry, T.Thirupathi Rao, A.Sailaja, Venkateshwara Rao; J, Indian Drugs, **28**, 489 (1991).

Regular Paper

- [25] S.M.EL-Ashry, F.A.Irbahim; *Anal.Lett*, **25**, 1657 (1992).
- [26] K.R.Mahadik, D.G.Rudraawas, H.N.More, S.S.Kadam; *Indian Drugs*, **28**, 530 (1991).
- [27] N.M.Sanghavi, M.M.Samarth, J.Warrier; *Indian Drugs*, **28**, 567 (1991).
- [28] T.Jovanovic, B.Stankovic, Z.Koricanac; *J Pharm Biomed Anal*, **13**, 213 (1995).
- [29] C.S.P.Sastry, K.R.Srinivas, K.M.Prasad; *Anal Lett*, **29**, 1329 (1996).
- [30] C.S.P.Sastry, S.G.Rao, P.Y.Naidu, K.R.Srinivas; *Anal Lett*, **31**, 263 (1998).
- [31] K.M.Emara, A.M.I.Mohamad, H.F.Askal, I.A.Darwish; *Anal Lett*, **26**, 2385 (1993).
- [32] A.I.Vogel; *Text Book of Quantitative inorganic analysis*, edn. **3**, (ELBS, Longman, London), 349 (1978).
- [33] B.Kratochiril, M.C.White; *Anal Chim Acta*, **31**, 528 (1964).
- [34] F.Mohr; *Ann.*, **105**, 60 (1958); E.cf Muller, O.Diefenthaler; *Z.Anorg; Chem*, **67**, 418 (1910); M.Kohn; *Anal.Acta*, **10**, 405 (1954).
- [35] U.Muralikrishna, A.Shiva Ramakrishna, T.A.Santhishree, S.S.Boce; *Indian Council of Chemists, 16th Conference* 159 AO-17 P (1997).